A forward genetic screen identifies modifiers of a voltage- and calcium-activated K⁺ channel in left-right neuronal asymmetry

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The developing nervous system generates a large diversity of cell types with distinct patterns of gene expression and functions. One way to establish neuronal diversity is to specify neuronal subtypes across the left-right axis. The *C. elegans* left and right AWC olfactory neurons communicate to specify asymmetric subtypes, AWC^{OFF} and AWC^{ON}. The default AWC^{OFF} is specified by a Ca^{2+} -regulated kinase cascade that is activated by influx of Ca²⁺ through the voltage-gated Ca²⁺ channel UNC-2/UNC-36. Intercellular communication between the two AWC neurons and other neurons through the NSY-5/innexin gap junction network antagonizes unc-2/unc-36 Ca²⁺ signaling in the induced AWC^{ON} cell. Our recent data suggest that voltage- and calcium-activated SLO BK potassium channels *slo-1* and *slo-2* acts redundantly downstream of *nsy-5* to inhibit *unc-* $2/unc-36 \text{ Ca}^{2+}$ signaling in the specification of AWC^{ON}. To identify the genes required for slo-1 function in inhibiting unc-2/unc-36 Ca²⁺ signaling for promoting AWC^{ON}, we performed a non-biased forward genetic screen to isolate *mok* (modifier of K⁺ channel) mutants that suppress the *slo-1(gf)* 2-AWC^{ON}-neuron phenotype. From about 6,000 genomes screened, we identified 16 new mutants that define genes required for *slo-1* function in promoting AWC^{ON}. The molecular lesions of all these mok mutants were identified using one-step whole genome sequencing and SNP mapping. Molecular characterization of these mok genes will begin to address how gap junction-mediated transient signaling coordinates long-term stochastic neuronal subtypes through downstream Ca2+-activated K+ channels and MOK molecules. Strategies of forward genetic screens and one-step whole genome sequencing/SNP mapping will be presented in the seminar and workshop.